

**Product Cat. No.: FP-251**

**For Clinical Diagnosis & Scientific Research.**

## FGFR2 (10q26) gene break apart probe reagent Instructions Manual

**[Product Name]** FGFR2 (10q26) gene break apart probe reagent.

**[Package Specifications]** 10 Tests/box.

### **[Intended use]**

In situ hybridization (ISH) was performed on the basis of routine staining to provide auxiliary information for diagnosis. The test results are only for clinical reference and should not be used as the only basis for clinical diagnosis. Clinicians should make a comprehensive judgment on the test results in combination with the patient's condition, drug indications, treatment response and other laboratory test indicators.

### **[Detection principle]**

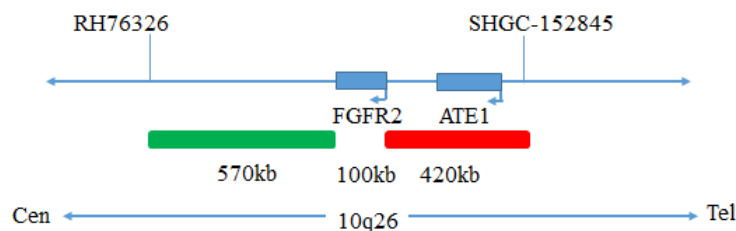
Fluorescence in situ hybridization (FISH) is a technique to directly observe specific nucleic acids in cells in vitro. According to the principle of base complementary pairing, a specific probe is complementary to the target sequence in the cell. Because the probe has fluorescence, the hybridization probe and the target sequence can be clearly observed under the fluorescence microscope under the appropriate excitation light.

### **[Product Main Components]**

The kit consists of FGFR2 dual color probe, as shown in Table 1.

**Table 1: Kit composition**

Component name	Specifications	Quantity	Main components
FGFR2 dual color probe	100μL/Tube	1	FGFR2 orange probe ; FGFR2 green probe



### **[Storage conditions & Validity]**

Keep sealed away from light at -20°C±5°C. The product is valid for 12 months. Avoid unnecessary repeated freezing and thawing that should not exceed 10 times. After opening, within 24 hours for short-term preservation, keep sealed at 2~8°C in dark. For long-term preservation after opening, keep the lid sealed at -20°C±5°C away from light. The kit was transported below 0°C.

### **[Applicable Instruments]**

Fluorescence microscope imaging system, including fluorescence microscope and filter set suitable for DAPI (367/452), green (495/517) and orange (547/565).

### **[Sample Requirements]**

1. Applicable specimen type: paraffin embedded specimen of surgical resection or biopsy tissue.

2. The tissue in vitro should be fixed with 4% neutral formaldehyde fixative within 1 hour. After the tissue is fixed, it is often dehydrated and embedded in paraffin.

#### [Testing Method]

##### 1. Hybridization pretreatment

Baking: Slides heating at 80°C for 30min or 65°C for 2h or overnight.

Dewaxing: According to the customer laboratory protocol (Commonly with Xylene for 15min).

Hydration: Take out the slides and put them respectively into 100%, 85% and 70% EtOH at room temperature for 3 minutes each.

Take out the slides, and immerse them in deionized water for 3 minutes. Remove the excess of water on the slides by air-drying.

Permeation: Immerse the slides in deionized water at 100°C and boil continuously for 20-40 minutes (Conventional 20min). Remove the excess of water on the slides by air-drying.

Digestion: Protease enzymic digestion at 37°C for 10-40 minutes. Mix the protease work buffer (50mmol HCl) and the 10x protease solution (Pepsin concentration 5%) in a proportion of 9:1 to prepare the enzymatic digestion solution.

Washing: Wash with 2xSSC at room temperature for 5 minutes.

Dehydration: Take out the slides and dehydrate in 70%, 85%, and 100% gradient ethanol at room temperature for 2 minutes each time. Remove the excess of EtOH solution on the slides by air-drying.

##### 2. Denaturation and Hybridization

The following operations should be carried out in the dark room.

① Take out the probe, let it stand at room temperature for 5min, turn it upside down with force, mix the probe well, centrifuge it briefly (do not vibrate with vortex apparatus), drop 10μl into the hybridization area of the cell drop, cover the 22mm×22mm cover glass immediately, the probe should be evenly spread under the cover glass without bubbles, and seal the edge with rubber (the edge sealing must be thorough to prevent the dry slide from affecting the test results in the hybridization process).

② Place the glass slide in the hybridization instrument, denature at 85°C for 5 min (the hybridizer should be preheated to 85°C) and hybridized at 42°C for 2-16h.

##### 3. Washing

The following operations should be carried out in a dark room.

① Carefully tear off the adhesive around the cover glass with tweezers to avoid sticking off or moving the cover glass. Immerse the cell drop into 2xSSC for about 5s, and take it out. Gently push one corner of the cover glass to the edge of the slide with tweezers, and gently remove the cover glass with tweezers;

② The cells were placed at 2xSSC room temperature for 1min;

③ Take out the slides and immerse in a preheated at 68°C 0.3% NP-40/0.4xSSC (Preparation of 0.3% NP-40/0.4xSSC: For 1L preparation, take 3mL NP-40 and 20mL 20xSSC, dissolve fully, mix well, and use 1M NaOH to adjust the pH to 7.2) solution and wash for 2min.

④ The slides were immersed in deionized water preheated at 37°C for 1min, and then dried naturally in the dark.

##### 4. Counterstaining

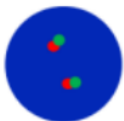

The following operations should be performed in a darkroom

10μl DAPI compound dye is dropped in the hybridization area of the glass slide and immediately covered. The suitable filter is selected for glass slide observation under the fluorescence microscope.

##### 5. FISH results observation

Place the stained sections under a fluorescence microscope and the cells area is first confirmed under a low magnification objective (10x); under magnification objective (40x) a uniform cells distribution is observed; then the nucleus size uniformity, nuclear boundary integrity, DAPI staining uniformity, no nuclei overlapping, cells clear signal are observed in the high magnification objective (100x).

#### [Interpretation of common signal types]

<p>● FGFR2 gene site 5 signal</p> <p>● FGFR2 gene site 3 signal</p>	
	Negative: 2 fusion
	Positive : 1 orange 1 green 1 fusion

#### [Limitations of test methods]

- ① The results of this kit will be affected by various factors of the sample itself, but also limited by hybridization temperature and time, operating environment and the limitations of current molecular biology technology, which may lead to wrong results.
- ② Users must be aware of the potential errors that may exist in the detection process and the limitations of accuracy.

#### [Precautions]

1. Please read this manual carefully before testing. The testing personnel shall receive professional technical training, and the signal counting personnel must be able to observe and distinguish orange and green signals.
2. When testing clinical samples, when the hybridization signal counting is difficult and the sample is not enough to repeat the retest, or the cell volume is not enough for analysis, the test will not provide the test results.
3. DAPI counterstaining agent used in this experiment has potential toxicity or carcinogenicity, so it is necessary to operate in the fume hood, wear masks and gloves to avoid direct contact.
4. All chemicals are potentially dangerous. Avoid direct contact. Used kits are waste and should be properly disposed off.

#### [Basic information]

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#### [Manual Approval date & Revision date]

V1. 0: Approval date: November 4, 2019.

V1. 2: Revision date: December 7, 2021.